

# MaxQuant data analysis in Galaxy

# Schedule

- Demo: Using MaxQuant in Galaxy
- **Hands-on:** Label-free proteomics analysis from skin cancer samples

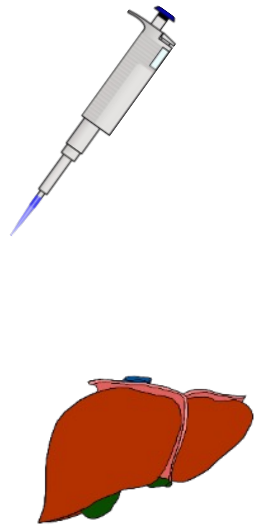
# Hands-on analysis / homework

- Tutorial: MaxQuant and Msstats for the analysis of label-free data  
<https://training.galaxyproject.org/training-material/topics/proteomics/tutorials/maxquant-label-free/tutorial.html>
- Share the Galaxy History:
  - Gear wheel on the upper right corner of the history
  - share or publish
  - Enable: make history accessible
  - Copy the link that is shown below

# Sample preparation

# Mass spectrometry

# Data analysis



Protein extraction



Protein(s)



Reduction & Alkylation



Linear Protein(s)



Tryptic digestion



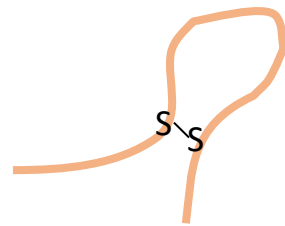
Peptides



Desalting  
Drying



Peptide Pellet



DAHSFGBKDFBVKNXSRMGLX

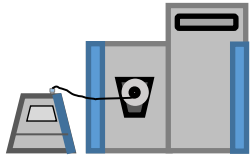


DAHSFGBK  
DFBVKNXSR  
MGLX

Sample preparation

Mass spectrometry

Data analysis



Peptide  
Pellet



Solving in  
acidic buffer



Liquid  
Chromato-  
graphy (LC)



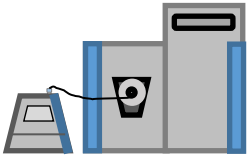
Ion  
source



Mass  
Analyzer



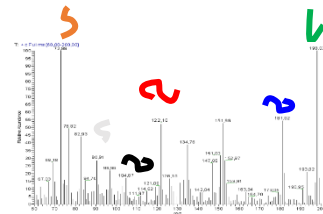
Detector



Tandem mass spectrometry (LC-MS/MS)  
using data dependent acquisition (Top3)

LC + Ion  
source

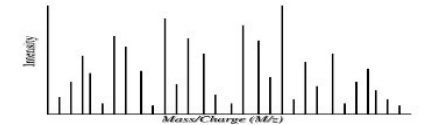
MS1: Mass spectra



Filter for most  
abundant m/z

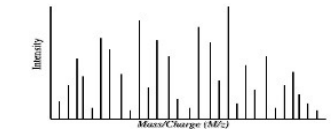
Fragmentation  
of filtered m/z

MS2: Mass spectra



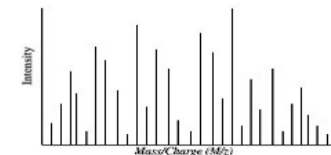
Filter for 2<sup>nd</sup> most  
abundant m/z

Fragmentation  
of filtered m/z



Filter for 3<sup>rd</sup> most  
abundant m/z

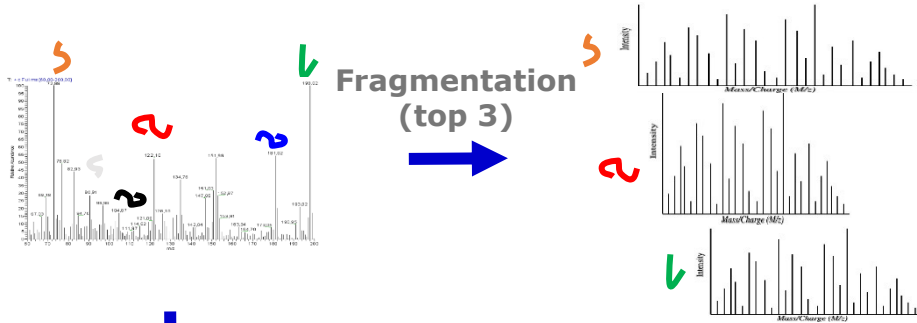
Fragmentation  
of filtered m/z



Next MS1 spectrum etc...

MS1: Mass spectra

MS2: Mass spectra



Peptide  
quantification

Peptide  
identification

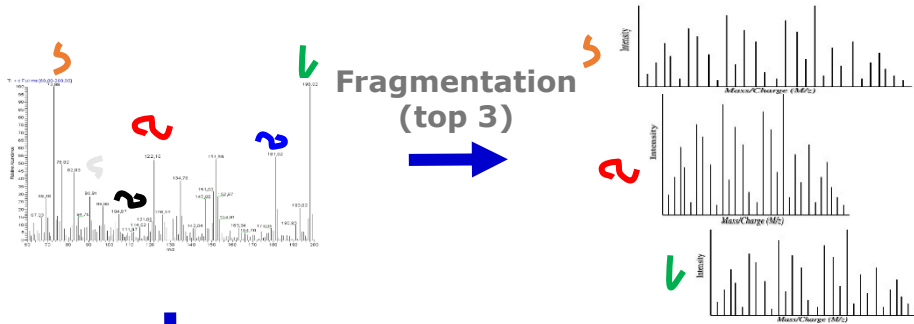
Sample preparation

Mass spectrometry

Data analysis

MS1: Mass spectra

MS2: Mass spectra



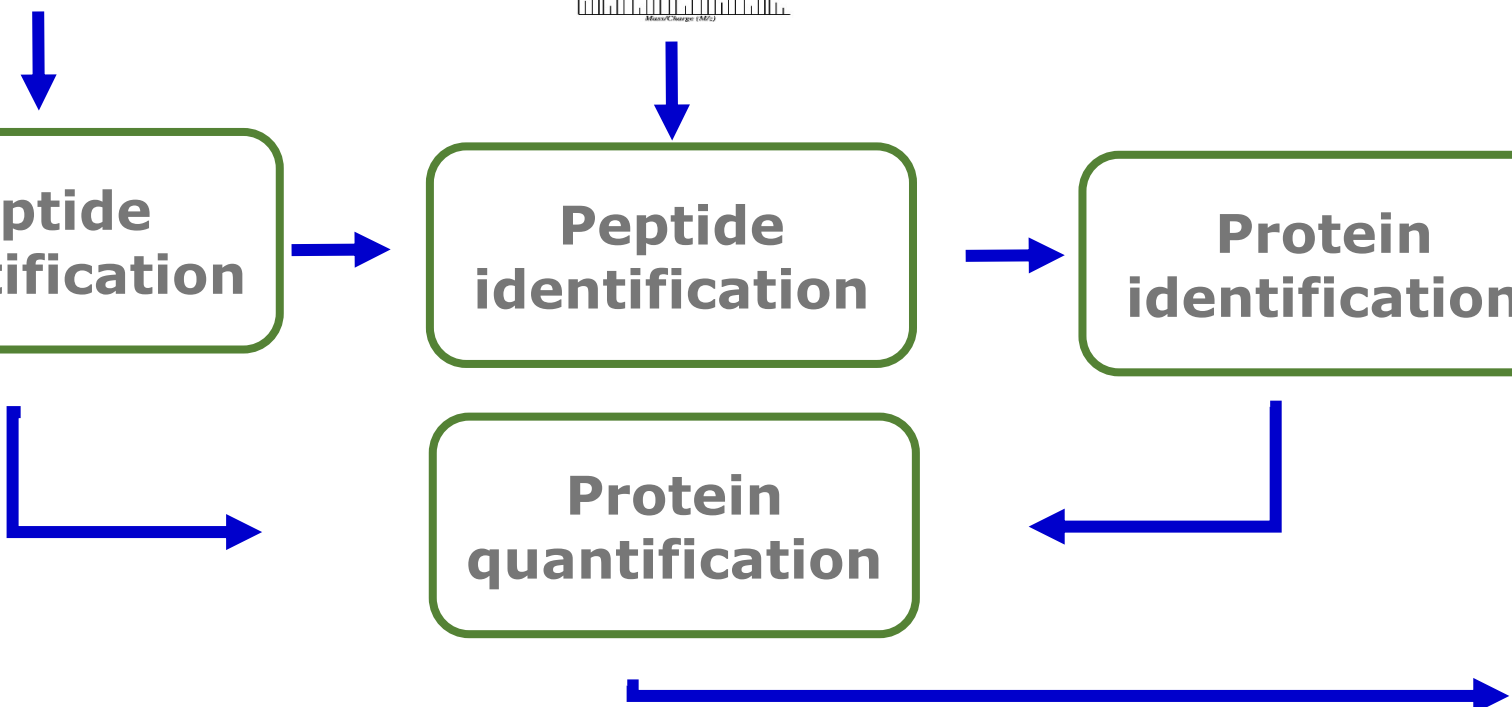
Peptide  
quantification

Peptide  
identification

Protein  
identification

Protein  
quantification

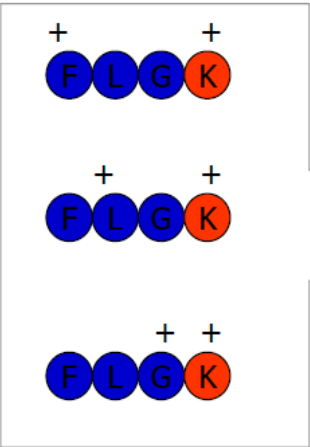
Statistical  
Analysis





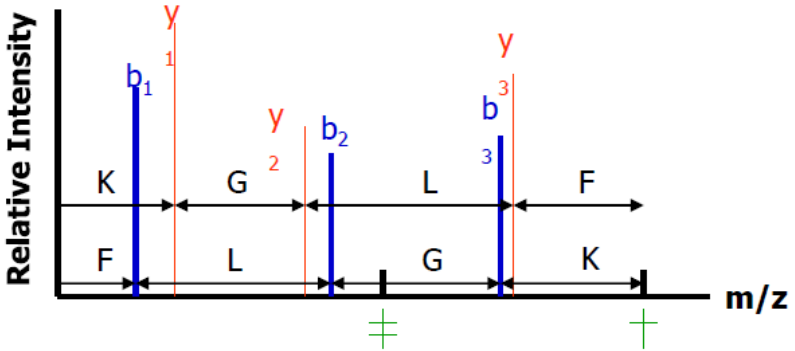
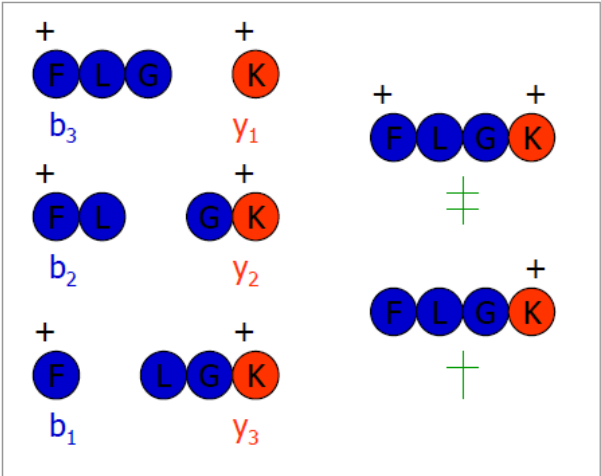
# Peptide identification with MS2 fragment spectra

MS1 spectra:  
Precursor ions



CID

MS2 spectra:  
Fragment ions



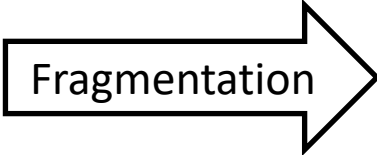
Identification options:

- 1) Manual interpretation
- 2) De novo sequencing
- 3) Matching to in silico spectra (generated from fasta database)

# Peptide identification with MS2 fragment spectra

MS1 spectra: Precursor ions

V-H-L-T-P-E-E-K



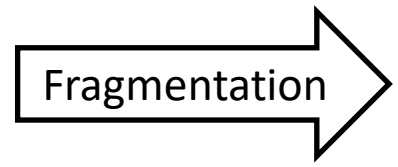
MS2 spectra: Precursor ions

	b-ions	y-ions	
b7	V-H-L-T-P-E-E	K	y1
b6	V-H-L-T-P-E	E-K	y2
b5	V-H-L-T-P	E-E-K	y3
b4	V-H-L-T	P-E-E-K	y4
b3	V-H-L	T-P-E-E-K	y5
b2	V-H	L-T-P-E-E-K	y6
b1	V	H-L-T-P-E-E-K	y7

# Peptide identification with MS2 fragment spectra

MS1 spectra: Precursor ions

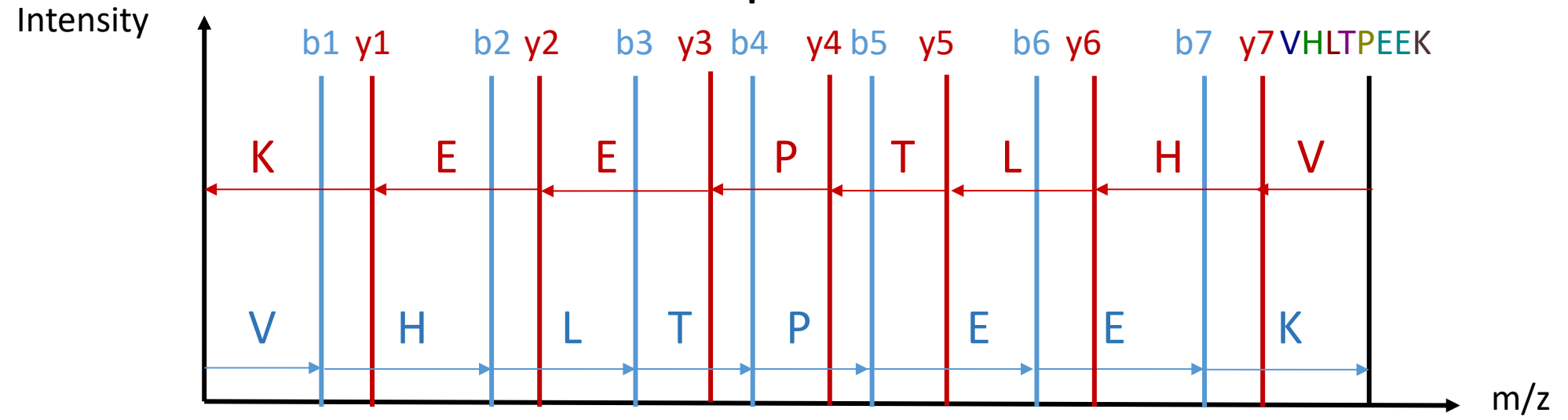
V-H-L-T-P-E-E-K



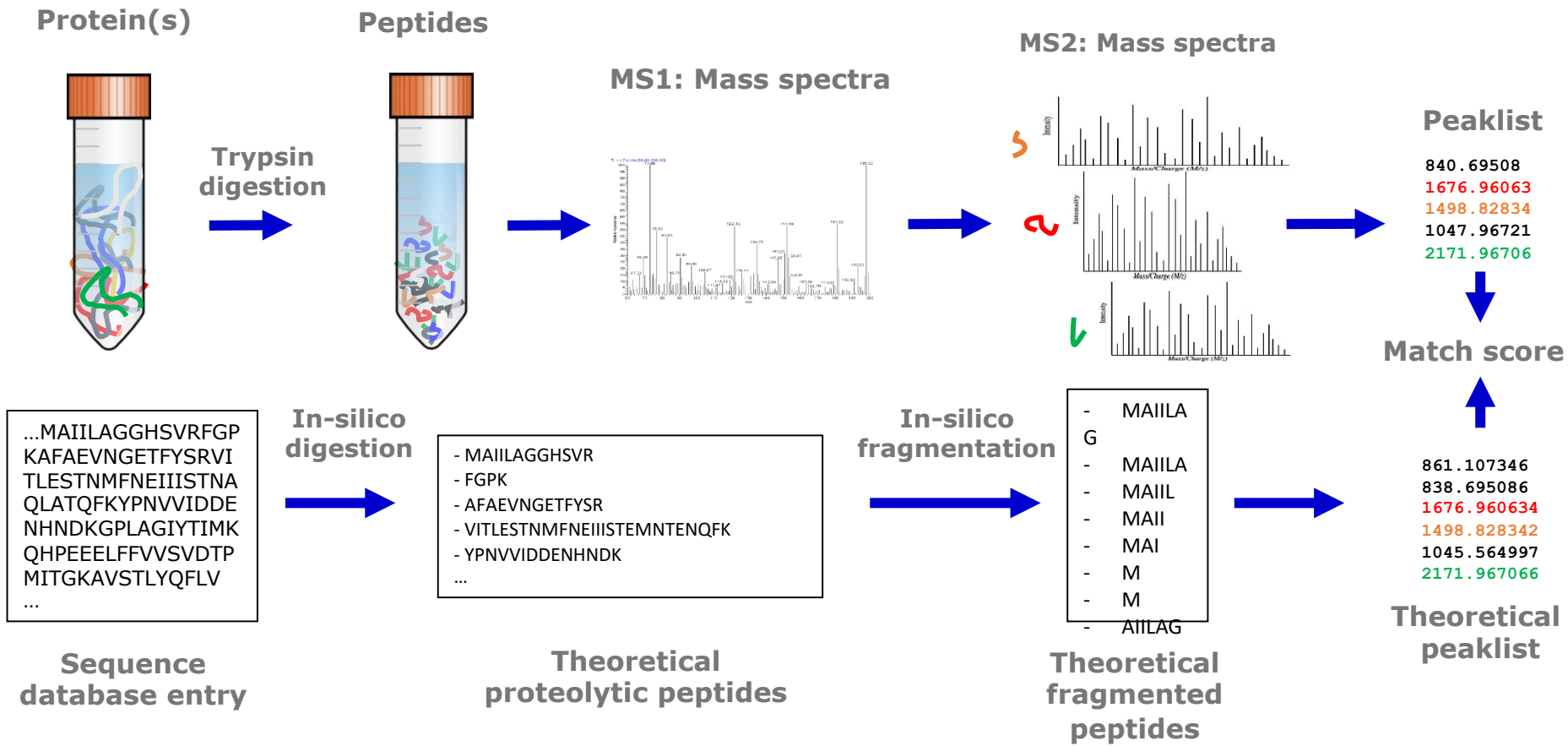
MS2 spectra: Precursor ions

b7	V-H-L-T-P-E-E	K	y1
b6	V-H-L-T-P-E	E-K	y2
b5	V-H-L-T-P	E-E-K	y3
b4	V-H-L-T	P-E-E-K	y4
b3	V-H-L	T-P-E-E-K	y5
b2	V-H	L-T-P-E-E-K	y6
b1	V	H-L-T-P-E-E-K	y7

MS2 spectrum



# Peptide identification via an in-silico database



...MAIILAGGHSVRFGP  
 KFAEVNGETFYSRVI  
 TLESTNMFNEIIISTNA  
 QLATQFKYPNVVIDDE  
 NHNDKGPLAGIYTIMK  
 QHPHEELFFVVSVDTP  
 MITGKAVSTLYQFLV  
 ...

- MAIILAGGHSVR  
 - FGPK  
 - AFAEVNGETFYSR  
 - VITLESTNMFNEIIISTEMNTENQFK  
 - YPNVVIDDENHNDK  
 ...

- MAIILA  
 G  
 - MAIILA  
 - MAIIL  
 - MAII  
 - MAI  
 - M  
 - M  
 - AIILAG

**Peaklist**

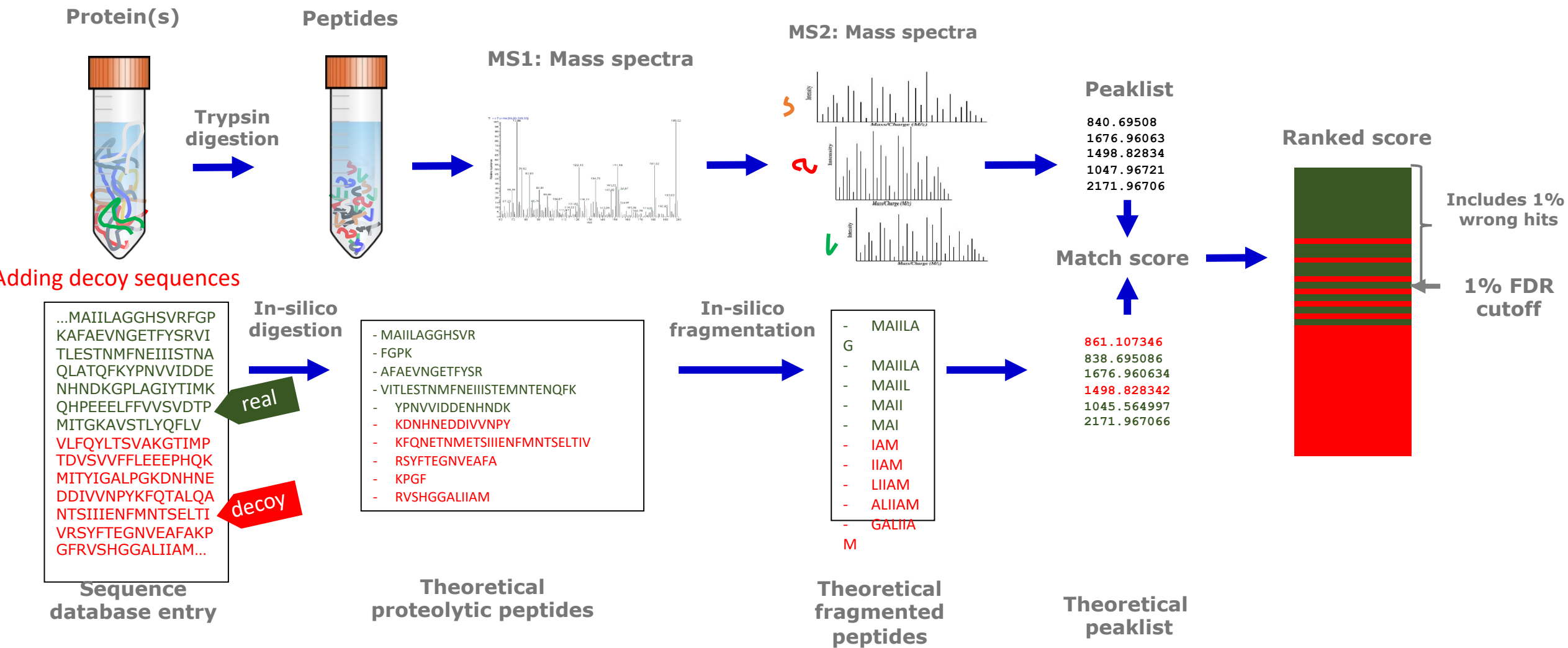
840.69508  
 1676.96063  
 1498.82834  
 1047.96721  
 2171.96706

**Match score**

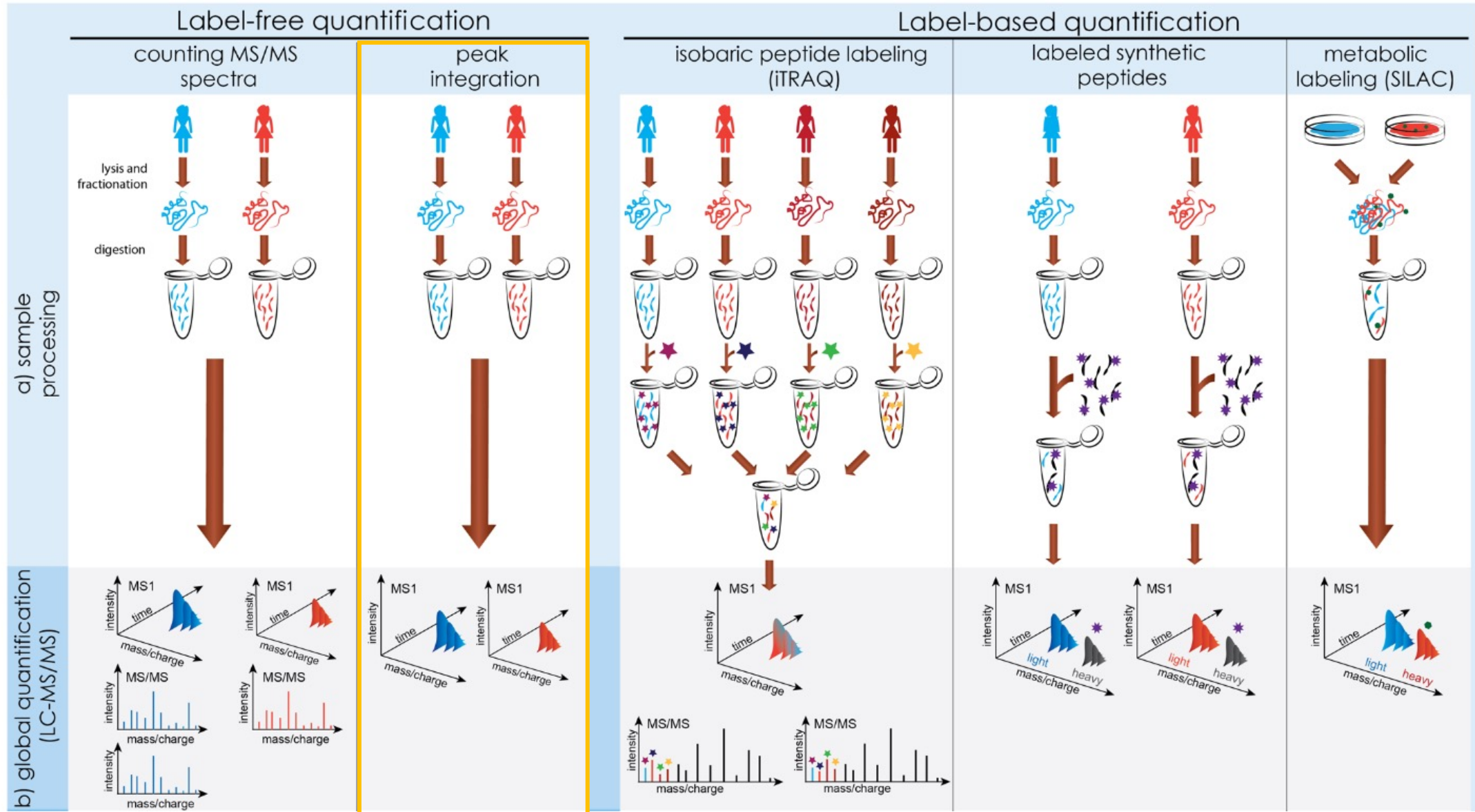
861.107346  
 838.695086  
 1676.960634  
 1498.828342  
 1045.564997  
 2171.967066

**Theoretical peaklist**

# Peptide identification via an in-silico database

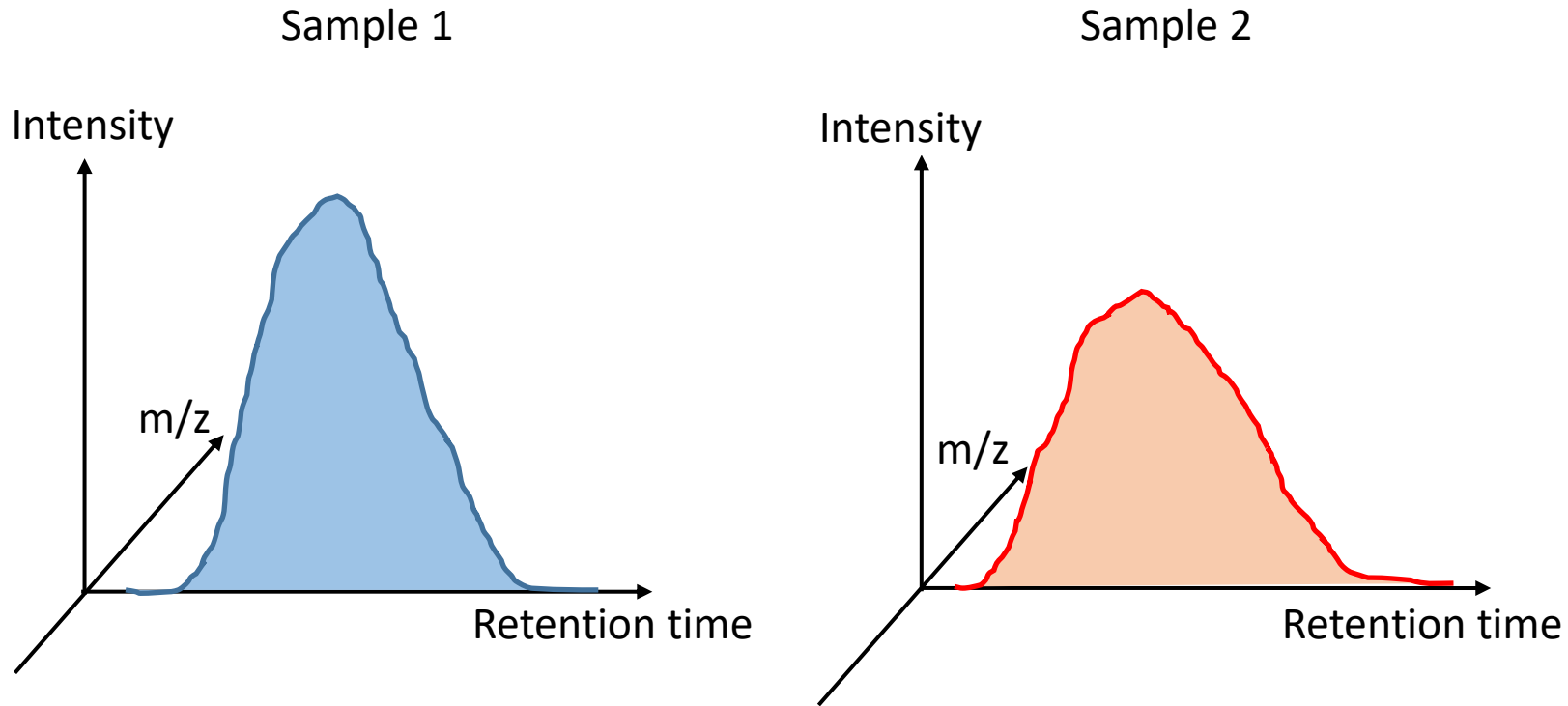


# Quantification methods in proteomics



# Label-free peptide quantification

MS1 spectra: for each peptide feature peak quantify area under curve to obtain peptide abundance



# MaxQuant software

Published: 30 November 2008

## **MaxQuant enables high peptide identification rates, individualized p.p.b.-range mass accuracies and proteome-wide protein quantification**

Jürgen Cox  & Matthias Mann 

*Nature Biotechnology* **26**, 1367–1372(2008) | [Cite this article](#)

**12k** Accesses | **6421** Citations | **12** Altmetric | [Metrics](#)

Freeware, “Black box”

Popular non-commercial proteomics software

MaxQuant videos on youtube:

<https://www.youtube.com/channel/UCKYzYTm1cnmc0CFAMhxDO8w>

- **Raw data import**
- **Protein Identification:**
  - Andromeda Search Engine
  - Speciality: Match-between-runs
- **Protein Quantification:**
  - Label-free
  - Label-based (SILAC, Dimethyl, ...)
  - Reporter ion MS2 (TMT, iTRAQ)



# MSstats software



## MSstats: an R package for statistical analysis of quantitative mass spectrometry-based proteomic experiments FREE

Meena Choi, Ching-Yun Chang, Timothy Clough, Daniel Broudy, Trevor Killeen, Brendan MacLean, Olga Vitek ✉ [Author Notes](#)

*Bioinformatics*, Volume 30, Issue 17, 1 September 2014, Pages 2524–2526, <https://doi.org/10.1093/bioinformatics/btu305>

**Published:** 02 May 2014 **Article history** ▼

Two Bioconductor R packages: Msstats & MSstatsTMT

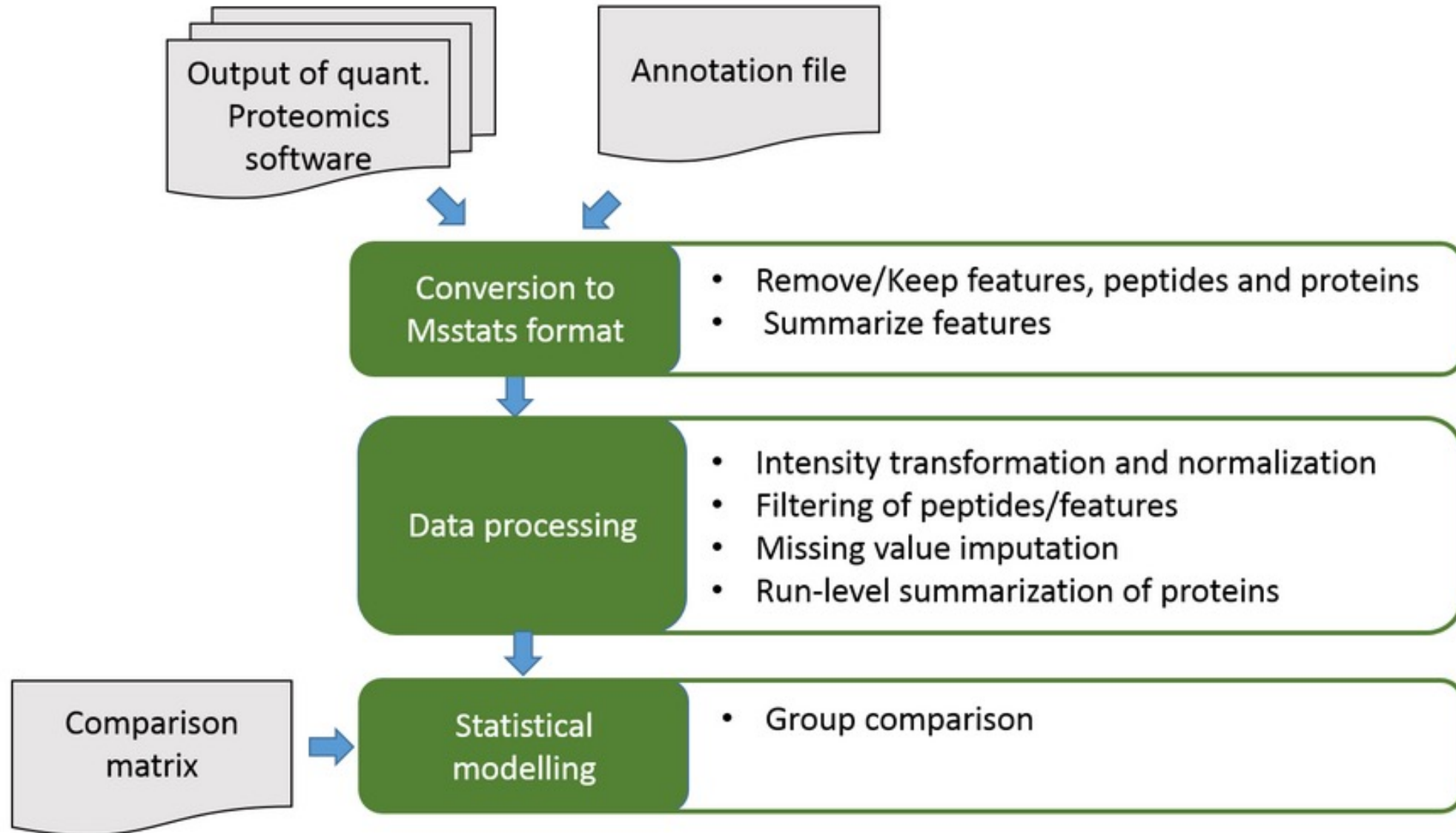
Popular open-source statistical proteomics software

MSstats videos on youtube:

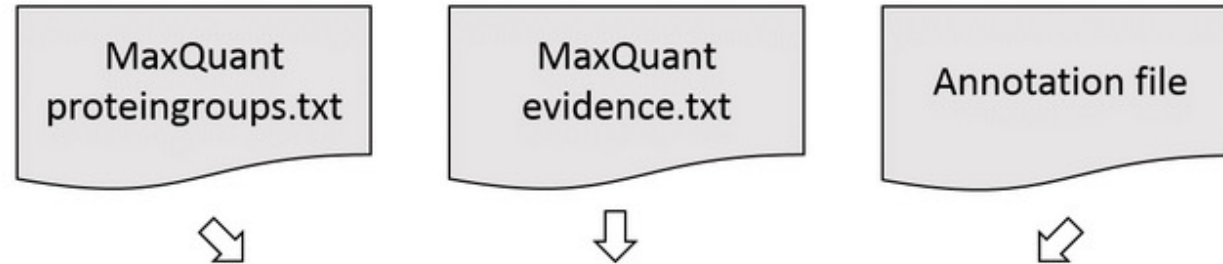
<https://www.youtube.com/c/MayInstituteNEU/search?query=msstats>

- **Import results from common proteomics software:**
  - MaxQuant, Skyline, OpenMS
- **Transformation and normalization**
- **Missing value imputation**
- **Protein summarization and quantification**
- **Statistical modelling**
  - linear models to detect differentially abundant proteins in label-free and isobaric labeled experiments

# Statistical Analysis with MSstats



# Conversion from MaxQuant to Msstats format



<b>Protein Name</b>	<b>Peptide Sequence</b>	<b>Precursor Charge</b>	<b>Fragm entlon</b>	<b>Product Charge</b>	<b>Isotope LabelType</b>	<b>Condition</b>	<b>Bio Replicate</b>	<b>Run</b>	<b>Intensity</b>
ProteinA	TPAVLK	3	NA	NA	L	Cond1	1	1	2636791
ProteinA	TPAVLK	3	NA	NA	L	Cond1	2	2	5019594
ProteinA	TPAVLK	3	NA	NA	L	Cond1	3	3	4560462
ProteinA	TPAVLK	3	NA	NA	L	Cond2	4	4	2918293
ProteinA	TPAVLK	3	NA	NA	L	Cond2	5	5	4534487

# Peptide filtering and missing value imputation

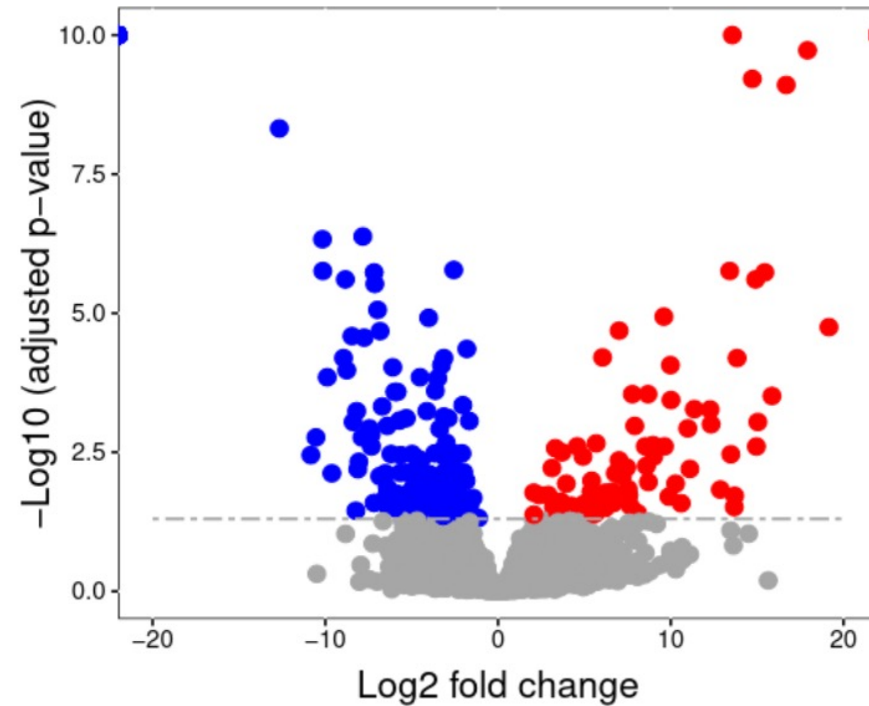
- **Feature selection:** which peptides should be kept for protein quantification
- **Missing value imputation:** Imputation of NA and very low abundant intensities
- **Protein summarization:** calculates new protein abundances after data processing via TMP model

	Condition1			Condition2		
	Subject1	Subject2	Subject3	Subject4	Subject5	Subject6
<b>Protein1</b>	Run1	Run2	Run3	Run4	Run5	Run6
Peptide1	Yellow	Grey	Grey	Grey	Light Yellow	Light Yellow
Peptide2	Orange	Grey	Light Yellow	Grey	Grey	Yellow
Peptide3	Orange	Red	Orange	Red	Orange	Orange
Peptide4	Grey	Yellow	Orange	Red	Yellow	Light Yellow
Peptide5	Red	Orange	Yellow	Orange	Grey	Grey

# Statistical modelling

- Uses run-level summarized data for hypothesis testing
- Needs comparison matrix to specify comparisons
- Adjusts the linear model according to information from annotation file

<b>name</b>	<b>Cond1</b>	<b>Cond2</b>	<b>Cond3</b>	<b>Cond4</b>
cond1-cond3	1	0	-1	0
cond1-cond4	1	0	0	-1
cond2-cond3	0	1	-1	0
cond2-cond4	0	1	0	-1





# Hands-on

Tutorial: MaxQuant and Msstats for the analysis of label-free data  
<https://training.galaxyproject.org/training-material/topics/proteomics/tutorials/maxquant-label-free/tutorial.html>

Video with demonstration of tutorial in youtube: <https://www.youtube.com/watch?v=IXdLAt2PAT4>

A few things might be outdated however, please stick to the recommendations of the training material which is up to date

In the first hands-on skip step 4 to 7 (don't load the raw data from PRIDE)  
Instead of running MaxQuant, load the MaxQuant result files from Zenodo.  
The links can be obtained by opening the box:  
Continue with the hands-on: MSstats Analysis

Tip: Continue with results from Zenodo

Because the MaxQuant run takes really long, we recommend to download the MaxQuant results from Zenodo and continue with the tutorial

1. Import the files from Zenodo

```
https://zenodo.org/record/4896554/files/MaxQuant_Evidence.tabular  
https://zenodo.org/record/4896554/files/MaxQuant_proteingroups.tabular  
https://zenodo.org/record/4896554/files/PTXQC_report.pdf
```

**Aim:** Find proteins that are differentially abundant between two different types of skin cancer:  
metastasizing cutaneous squamous cell carcinoma (cSCC) and recessive dystrophic epidermolysis bullosa cSCC

Metastasizing cSCC  
(n=13)

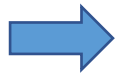


RDEB cSCC (n=6)

Per patient: 1-2 10µm slices  
of FFPE tissues

# HANDS-ON SESSION

## Instructions



Please **Register** for creating an account with a valid email ID and Password at [usegalaxy.eu](https://usegalaxy.eu).

Once Registered, click on TIAAS to join the GCC 2022 Galaxy session.  
<https://usegalaxy.eu/join-training/quant-meta>

Go to Shared Data

Go to Shared Data

Run the workflow on active history



# Further resources

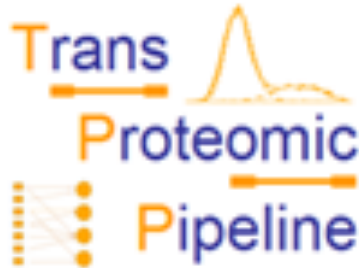
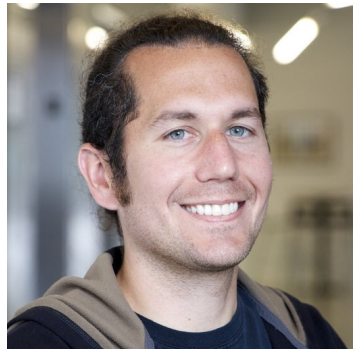
- Summary of proteomics data analysis:  
<https://www.youtube.com/watch?v=2C96AvrFT38>
- More proteomics tutorials in Galaxy:  
<https://training.galaxyproject.org/training-material/topics/proteomics>
- UC Davis Proteomics:  
[https://video.ucdavis.edu/playlist/details/0\\_4jkc4swu](https://video.ucdavis.edu/playlist/details/0_4jkc4swu)
- Global online Galaxy course in March (much more than proteomics)  
<https://gallantries.github.io/posts/2021/12/14/smorgasbord2-tapas/>



# Cloud Computing Workshop (2022)

The iPRG will conduct a series of online video tutorials about the use of cloud computing resources for MS-based proteomics, focusing on Nextflow, the Trans-Proteomic Pipeline (TPP) and Galaxy Platform.

**September 2022**



Michael Hoopmann - *ISB, Seattle, WA*

- Instructions on how to use TPP to analyze MS data.
- Answer questions from the participants

**October 2022**



Melanie Foell - *Freiburg University, Germany*

- Instructions on how to use Galaxy to analyze MS data.
- Answer questions from the participants

**November 2022**



Yasset Perez-Riverol - *EBI, Hinxton, UK*

- Instructions on how to use Nextflow to analyze MS data.
- Answer questions from the participants